



Synthesis and Biological Evaluation of a Conformationally Free *seco*-Analogue of the Immunosuppressant FR901483

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Received 17 June 1999; accepted 19 July 1999

Abstract—The synthesis of an azaspirocyclic analogue of FR901483, phosphate **2**, is described based on the implementation of a key 5-endo aminocyclization promoted by iodine for direct functionalization of the 1-azaspiro[4.5]decane ring at the C-3 atom. Compound **2** has no inhibitory activity in the cell proliferation assays reported. © 1999 Elsevier Science Ltd. All rights reserved.

Introduction

FR901483 (1) is a novel immunosuppressant recently isolated from the fermentation broth of *Cladobotryum* sp. No. 11231 and characterized by the Fujisawa Laboratories.¹ Immunosupressive agents display a wide variety of structures,^{2,3} but none are related to that of 1. This compound has a tricyclic ring system unprecedented among natural products, consisting of a 2-azabicyclo[3.3.1]nonane nucleus fused to a pyrrolidine moiety, resulting in a spiro framework. Additionally, 1 has a phosphate unit, which plays an important role in this compound's immunosuppressive activity. Recently, the unique structure of FR901483 has been the subject of two synthetic approaches directed to its tricyclic ring system,^{4,5} the Snider approach⁵ reaching the demethylamino derivative of 1.

Part of our ongoing research project towards the synthesis of FR901483 (1) is concerned with simplifying the structure and increasing the flexibility of the molecule; this has led to the consideration of the spiro derivative 2 (R=Me, Fig. 1). The *seco*-analogue 2 has a conformational mobility that 1 lacks, and constitutes an interesting target for pharmacological evaluation. Considering the chair axial—axial conformation for compound 2, a good overlap between pyrrolidine and methylamino nitrogen, as well as for the phosphate unit,

Key words: Immunological activity; natural products; phosphonic acid and derivatives; poliycyclic heterocyclic compounds.

can be seen in the superimposition showed in Figure 2. However, it is worth pointing out the absence in 2 of the hydroxyl group present in FR901483 and the slight separation of the methoxyphenyl moieties.

In this paper, we describe the synthesis and biological evaluation of the *seco*-analogue **2** (R=Bn),⁶ introducing a new synthetic procedure to obtain its azabicyclyc framework.

Results and Discussion

Synthesis

Our procedure for the synthesis of 1-azaspiro-[4.5]decanes (for other approaches leading to this azabicyclic system by N(1)–C(2) bond formation in the last step, see refs 7–12; for other synthetic entries, starting from carbocyclic compounds, see inter alia, refs 13-20) consists of the treatment of a homoallylamine (i.e. 3) with iodine (for successful examples of the scarce reactions of amino cyclization promoted by iodine, see refs 21 and 22; for iodine initiated cyclization of unsaturated nitrogen containing compounds other than amines, see refs 23-25), which promotes the electrophilic cyclofunctionalization of the unsaturated amine to give a pyrrolidine ring.²⁶ The required homoallylamine was prepared in a one-pot procedure involving the formation of an imine from the monoethylene acetal of 1,4-cyclohexanedione and 4-methoxyphenylethylamine followed by the addition of allylmagnesium bromide (Scheme 1).²⁷ Treatment of 3 with iodine promoted the

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Figure 1.

iodoaminocyclization process to give **4**. Thus, in only two separate steps we gained access to the suitably functionalized azabicyclic system. The oxidation of the iodo derivative **4** to ketone **5** was effected using DMSO and silver tetrafluoroborate. The overall sequence worked well and we obtained azaspiro **5** in 30% overall yield. Compound **5** was analyzed spectroscopically by means of NOESY experiments in order to elucidate its preferred conformation, which proved to be that where the amino group was disposed equatorially from the carbocyclic ring.

Having established a simple and efficient synthetic entry to 3,8-functionalized 1-azaspiro[4.5]decanes,²⁹ we next turned our attention to the preparation of the *seco*-analogue of FR901483 **2** (R=Bn). The subsequent steps involved several functional transformations: (i) reductive amination to introduce the amino group at C-3; (ii) deprotection of the acetal group in **6**, followed by

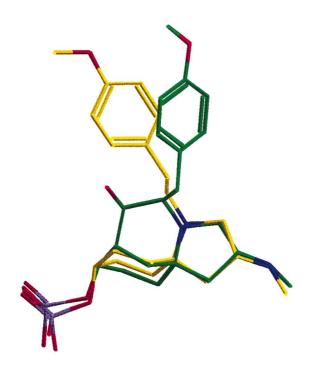


Figure 2. Molecular fitting of FR901483 (green) and 2 (yellow).

reduction of the resulting ketone 7 to stereoselectively afford the alcohol 8; (iii) protection of the secondary amino group, in order to avoid an oxidative process in later steps; (iv) assembly of the phosphomonoester group, which was achieved by phosphorylation of the hydroxyl group of 9 with O,O-bisbenzyl-N,N-diisopropyl phosphoramidate in the presence of tetrazole followed by MCPBA oxidation;³⁰ (v) finally, the benzyl phosphates of 10 were hydrogenolyzed and the resulting monoester 11 was treated with TFA to afford the diamino derivative 2, which was isolated as its dipotassium salt. The relative configuration at C-5 and C-8, which was generated in the ketone reduction step, was inferred taking into account the preferred conformation of ketone 7 (the same as that of the corresponding related acetal 5), and the fact that the NaBH4 reduction furnished an equatorial alcohol, as expected. Additionally, NOESY experiments upon 2 agreed with the configuration and conformation depicted in Scheme 1.

Biological evaluation

The synthesized simplified analogues of 1 (compounds 2, 7 and 8) were tested in the three lymphocyte in vitro proliferation assays indicated in Experimental (TPA plus IL-2, Tetanus toxoid or Phytohemegglutinin). None of these three compounds showed inhibitory activity at the range dose of 40 ng/mL to 4 mg/mL. The reference compound cyclosporin A showed good activity at 1 mg/mL.

Conformational studies

This lack of activity in the proliferation assay studies, particularly for compound 2 which is the closest analogue of 1, may be due to either the differences in functional groups between the compounds (absence of OH, benzyl in place of methyl) or an unsuitable conformation of 2, as suggested by the NMR experiments.

For a better understanding of this second possibility, a conformational analysis of the cyclohexane ring in the spiro moiety was carried out. We elected to evaluate this point by calculating the differences in the heat of formation for the two chair conformations. Comparisons of the resultant energy minima revealed that the conformation 2-I is indeed favored thermodynamically visà-vis the conformer 2-II; the corresponding heats of formation differed by 3.2 kcal/mol (Fig. 3). Thus, as suggested by the NMR experiments, the only conformation detected for compound 2 was that corresponding to a chair equatorial-equatorial (conformation 2-I) since the equilibrium constant between the two conformers is about 1000 at room temperature.³¹ This may suggest that one of the reasons for the lack of activity for compound 2 is the inaccessibility of the correct chair conformation.

Conclusions

Although compound 2 could in principle adopt a conformation that matches to a great extention that of

Scheme 1. Reagents and conditions: (i) 4-methoxyphenethylamine, molecular sieves; then allylmagnesium bromide, Et₂O:CH₂Cl₂, 71%; (ii) I₂, NaHCO₃, CH₂Cl₂:H₂O, 51%; (iii) AgBF₄, DMSO, 80%; (iv) benzylamine, molecular sieves; then NaBH₄ (85%); (v) HCl, THF:H₂O, 79%; (vi) NaBH₄, MeOH, 75%; (vii) (Boc)₂O. MeOH:Et₃N, 76%; (viii) (BnO)₂PN(*i*Pr)₂, tetrazole; then *m*-CPBA, CH₂Cl₂, -50°C, 73%; (ix), H₂, Pd/C, MeOH, 92%; (x) TFA, CH₂Cl₂; then K₂CO₃.

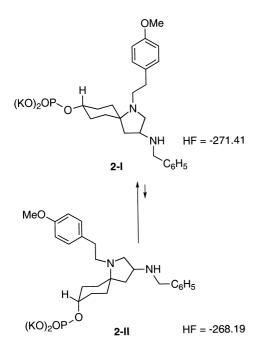


Figure 3. Conformational analysis of cyclohexane ring of compound **2** (HF = heat of formation in Kcal/mol, MOPAC/AM1).

structure 1 (see Fig. 1), in fact 2 adopts a *trans*-diequatorial conformational relationship between the phosphate group and the nitrogen atom, which is non-accessible to the natural product 1 in which both

substituents of the cyclohexane ring must be *trans*-diaxial for the additional ring linkage. As a consequence, the different distances between the various functional groups in **2** with respect to those in **1** may explain the lack of activity of the *seco*-analogue **2**.

From a synthetic standpoint, we report a new synthetic entry to the azaspiro[4,5]decane scaffold that might allow us to access the hitherto unprepared natural product FR901483, using an adequate amine (i.e. tryrosine derivative) as starting material which could allow the elaboration of the additional ring.

Experimental

Computational procedures

Structures were built with standard bond lengths and angles using the Chem-X^{32,33} molecular modelling package. All structures were initially optimised using the steepest descents and conjugate gradient methods. After semiempirical optimization using the MOPAC program,^{34,35} charge distributions were calculated by means of the AM1 method. The PULAY keyword and the eigenvector following the (EF) routine for minimum search with GNORM = 0.1 were used in the optimisation step. Conformation analysis was performed with fully AM1 optimization. All calculations were performed on a Digital Alpha Station 3000.

Biology

Assays were performed with freshly isolated human peripheral blood mononuclear cells isolated from blood by density gradient centrifugation. Lymphocyte proliferation induced by different stimulators (TPA plus IL-2, Tetanus toxoid or Phytohemagglutinin) were assessed by tritiated thymidine uptake. In all the cases 1×10^5 cells were placed in flat bottom wells from a microtiter plate in triplicates in a final volume of 200 mL of RPMI 1640 complete medium with or without test compound. For the TPA plus IL-2 induced proliferation assay nylon wool purified T cells were used. Cell proliferation was assessed on day 1 for TPA plus IL-2 and Phytohemagglutinin stimulation, whereas for Tetanus toxoid, induced proliferation was tested on day 6. Cyclosporin A was used as a control.

Chemistry

All reactions were carried out under an argon atmosphere with dry, freshly distilled solvents under anhydrous conditions. TLC was carried out on SiO₂ (silica gel 60 F₂₅₄, Merck), and the spots were located with UV light, iodoplatinate reagent or 1% aqueous KMnO₄. Chromatography refers to flash chromatography and was carried out on SiO₂ (silica gel, SDS, 230–400 mesh ASTM). Unless otherwise noted, drying of organic extracts during work up of reactions was performed over anhydrous Na₂SO₄. Evaporation of solvents was accomplished with a rotatory evaporator. ¹H NMR and ¹³C NMR spectra were recorded with a Varian 300 or a Varian VXR-500 instrument. Chemical shifts are reported in ppm downfield from Me₄Si. IR spectra were recorded on a Nicolet 205 FT-IR spectrophotometer, and only noteworthy absorptions are listed. Microanalyses were performed by the Centro de Investigación y Desarrollo (CSIC), Barcelona.

4-Allyl-4-[2-(4-methoxyphenyl)ethylaminolcyclohexanone ethylene acetal (3). To a solution of 1,4-cyclohexanedione monoethylene acetal (8 g, 51.2 mmol) in CH₂Cl₂ (160 mL) were added 2-(4-methoxyphenyl)ethylamine (9.8 mL, 66.6 mmol) and 4 Å molecular sieves (16 g). After stirring at room temperature for 4 h, the suspension was filtered through Celite, and the filtrate was concentrated to give the corresponding imine (13 C NMR ($^{74.5}$ MHz, CDCl₃) δ 24.3 (CH₂), 33.6 (CH₂), 34.5 (CH₂), 35.9 (CH₂), 36.0 (CH₂), 52.2 (CH₂N), 54.8 (CH₃O), 64.0 (CH₂O), 107.5 (C-1), 113.3 (CH), 129.4 (CH), 131.8 (C), 157.6 (C), 170.6 (CN)). To a solution of this imine in CH₂Cl₂ (80 mL) was added dropwise a solution of allyl magnesium bromide (102.4 mmol) in Et_2O (120 mL). The mixture was stirred at room temperature for 4h, poured into saturated aqueous NH₄Cl, and extracted with CH₂Cl₂. The organic extracts were washed with brine, dried, and concentrated. Chromatography (from CH₂Cl₂ to 9:1 CH₂Cl₂:MeOH) yielded amine 3 (12 g, 71%): ¹H NMR (500 MHz, CDCl₃) δ 0.75 (br, 1H, NH), 1.40–1.58 (m, 6H), 1.70–1.80 (m, 2H), 2.10 (d, J=7.5 Hz, 2H, $CH_2C=$), 2.66 (m, 4H, CH_2CH_2), 3.77 (s, 3H, OCH_3), 3.90 (m, 4H, CH₂O), 4.96 (ddt, J = 17, 2 and 1.5 Hz, 1H,

H-cis), 5.00 (ddt, J=10, 2.5 and 1.5 Hz, 1H, H-trans), 5.66 (ddt, J=17, 10 and 7.5 Hz, 1H, H-gem), 6.81 (d, J=8.5 Hz, 2H, 3'-H), 7.10 (d, J=8.5 Hz, 2H, 2'-H); ¹³C NMR (74.5 MHz, CDCl₃) δ 30.1 and 32.5 (C-2 and C-3), 36.2 (CH₂Ar), 41.4 (CH₂CH), 42.8 (CH₂N), 52.7 (C-4), 55.1 (CH₃O), 64.0 (CH₂O), 108.9 (C-1), 113.6 (C-3'), 117.5 (=CH₂), 129.5 (C-2'), 132.3 (C-1'), 134.0 (=CH), 157.8 (C-4'). Anal. calcd for C₂₀H₂₉NO₃: C, 72.47; H, 8.82; N, 4.23. Found: C, 72.26; H, 9.05; N, 4.30.

1-[2-(4-Methoxyphenyl)ethyl]-3-iodo-1-azaspiro[4.5]decan-8-one ethylene acetal (4). To a solution of amine 3 (2 g, 6 mmol) in CH₂Cl₂ (60 mL) and 5% aqueous NaHCO₃ (60 mL) was added dropwise a solution of I₂ (2.28 g, 9 mmol) in CH₂Cl₂ (60 mL). After stirring at room temperature for 12h, saturated aqueous sodium thiosulfate was added. The mixture was extracted with CH₂Cl₂. The dried organic extracts were concentrated and chromatographed (99.5:0.5 CH₂Cl₂:MeOH) to give compound 4 (1.41 g, 51%): ¹H NMR (300 MHz, CDCl₃) δ 1.36 (m, 1H), 1.45–1.85 (m, 7H), 2.26 (dd, J = 14 and 6.6 Hz, 1H, H-4), 2.50 (dd, J = 14 and 8.2 Hz, 1H, H-4), 2.69 (m, 4H, CH_2CH_2), 3.23 (dd, J=10.1 and 6.9 Hz, 1H, H-2), 3.39 (dd, J = 10.1 and 7.4 Hz, 1H, H-2), 3.79 (s, 3H, CH₃O), 3.91 (m, 4H, CH₂O), 4.27 (dddd, J = 8.2, 7.4, 6.9 and 6.6 Hz, 1H, H-3), 6.82 (d, J = 8.6 Hz, 2H, 3'-H), 7.12 (d, J = 8.6 Hz, 2H, 2'-H); ¹³C NMR (74.5 MHz, CDCl₃) δ 15.0 (C-3), 28.9 and 29.9 (C-6 and C-10), 31.9 and 32.4 (C-7 and C-9), 34.5 (CH₂Ar), 47.0 (C-4), 49.3 (CH₂N), 55.1 (CH₃O), 61.9 (C-2), 64.0 and 64.2 (CH₂O), 64.6 (C-5), 107.8 (C-8), 113.6 (C-3'), 129.6 (C-2'), 131.8 (C-1'), 157.8 (C-4'). Anal. calcd for C₂₀H₂₈INO₃: C, 52.52; H, 6.17; N, 3.06. Found: C, 52.76; H, 6.25; N, 3.02.

1-[2-(4-Methoxyphenyl)ethyl]-1-azaspiro[4.5]decan-3,8dione 8-monoethylene acetal (5). To a solution of AgBF₄ (0.68 g, 3.44 mmol) in DMSO (40 mL) was added dropwise a solution of 4 (1.64 g, 3.6 mmol) in DMSO (30 mL). After stirring at room temperature for 12 h, the suspension was filtered through Celite, and the filtrate partitioned between Et₂O and water. The organic extracts were washed with brine, dried, and concentrated. Chromatography (CH₂Cl₂:MeOH, 99:1) afforded ketone 5 (0.99 g, 80%): IR (film) 1758; ¹H NMR (500 MHz, CDCl₃) δ 1.37 (dm, J = 13.5 Hz, 2H, H-6eq and H-10eq), 1.50 (td, J = 13.5 and 4 Hz, 2H, H-7ax and H-9ax), 1.73 (dm, J = 13.5 Hz, 2H, H-7eq and H-9eq), 1.88 (td, J = 13.5 and 4 Hz, 2H, H-6ax and H-10ax), 2.37 (s, 2H, H-4), 2.65 (dd, J=9 and 6.5 Hz, 2H, CH_2Ar), 2.77 (dd, J = 9 and 6.5 Hz, 2H, CH_2N), 3.24 (s, 2H, H-2), 3.77 (s, 3H, CH₃O), 3.90 (s, 4H, CH₂O), 6.81 (d, J=9 Hz, 2H, 3'-H), 7.09 (d, J=9 Hz, 2H, 2'-H); ¹³C NMR (74.5 MHz, CDCl₃) δ 28.3 (C-6 and C-10), 32.4 (C-7 and C-9), 35.0 (CH₂Ar), 47.7 (C-4), 49.9 (CH₂N), 55.2 (CH₃O), 58.9 (C-2), 62.1 (C-5), 64.2 and 64.3 (CH₂O), 107.9 (C-8), 113.6 (C-3'), 129.6 (C-2'), 132.1 (C-1'), 157.9 (C-4'), 214.0 (C-3). Anal. calcd for C₂₀H₂₇NO₄: C, 69.54; H, 7.88; N, 4.05. Found: C, 69.59; H, 7.88; N, 4.24.

3-Benzylamino-1-[2-(4-methoxyphenyl)ethyl]-1-azaspiro-[4.5]decan-8-one ethylene acetal (6). To a solution of

ketone 5 (0.55 g, 1.6 mmol) in CH_2Cl_2 (2.8 mL) were added benzylamine (0.34 mL, 3.2 mmol) and 4 A molecular sieves (0.55 g). After stirring at room temperature for 4h, the suspension was filtered through Celite, and the filtrate was concentrated to give the corresponding imine. To a cooled (0°C) solution of this imine in MeOH (14 mL) was added NaBH₄ (121 mg, 3.2 mmol). After 2h at 0°C and 2h at room temperature, the reaction mixture was poured into saturated aqueous NH₄Cl and extracted with CH_2Cl_2 (5×50 mL). The organic extracts were dried and concentrated. Chromatography (CH₂Cl₂ to 9:1 CH₂Cl₂:MeOH) yielded amine 6 (0.59 g, 85%): ${}^{1}H$ NMR (500 MHz, CDCl₃) δ 1.20 (d, J = 9.5 Hz, 1H, H-6eq), 1.30 (d, J = 12.5 Hz, 1H, H-10eq), 1.46 (dd, J = 13 and 6 Hz, 1H, H-4), 1.49–1.66 (m, 5H, H-6ax, H-7 and H-9), 1.69 (td, J = 12.5 and 4 Hz, 1H, H-10ax), 2.12 (dd, J = 13 and 8.5 Hz, 1H, H-4), 2.51 (m, 1H, NCH₂), 2.56–2.68 (m, 3H, NCH₂ and CH₂Ar), 2.77 (dd, J = 9.5 and 4.5 Hz, 1H, H-2), 2.93 (dd, J = 9.5and 7 Hz, 1H, H-2), 3.24 (m, 1H, H-3), 3.68 (s, 2H, NCH₂Ar), 3.70 (s, 3H, CH₃O), 3.82 (m, 4H, OCH₂ CH_2O), 6.72 (d, J=8.5 Hz, 2H, 3'-H), 7.03 (d, J = 8.5 Hz, 2H, 2'-H), 7.17 (m, 1H), 7.21-7.28 (m, 4H);¹³C NMR (74.5 MHz, CDCl₃) δ 28.0 (C-6), 30.8 (C-10), 32.4 (C-7), 32.6 (C-9), 35.3 (CH₂Ar), 42.0 (C-4), 50.0 (NCH₂), 52.4 (NCH₂Ar), 54.5 (C-3), 55.0 (CH₃O), 57.2 (C-2), 62.5 (C-5), 64.1 and 64.2 (OCH₂CH₂O), 108.5 (C-8), 113.6 (C-3'), 126.9 (p-C), 128.2 and 128.4 (o-C and *m*-C), 129.6 (C-2'), 132.8 (C-1'), 140.2 (*ipso*-C), 157.8 (C-4'). Anal. calcd for C₂₇H₃₆N₂O₃.1/2H₂O: C, 72.77; H, 8.37; N, 6.29. Found: C, 73.08; H, 8.20; N, 6.69.

3-Benzylamino-1-[2-(4-methoxyphenyl)ethyl]-1-azaspiro-[4.5]decan-8-one (7). To a solution of acetal 6 (1.65 g, 3.8 mmol) in THF (30 mL) was added hydrochloric acid (10%, 83 mL). After 12h at room temperature, the reaction mixture was poured into 5% aqueous NaHCO₃ and extracted with CH_2Cl_2 (5×50 mL). The organic extracts were dried and concentrated. Chromatography (CH₂Cl₂ to 94:6 CH₂Cl₂:MeOH) afforded ketone 7 (1.18 g, 79%): IR (film) 1714; ¹H NMR (300 MHz, CDCl₃) δ 1.60 (br, 1H, H-6eq), 1.75 (m, 3H, H-4, H-6ax and H-10eq), 1.86 (td, J=13 and 5.5 Hz, 1H, H-10ax), 2.30-2.42 (m, 5H, H-4, H-7 and H-9), 2.56 (m, 1H, CH₂N), 2.62–2.72 (m, 3H, CH₂N and CH₂Ar), 2.89 (dd, J=9.5 and 4.5 Hz, 1H, H-2), 3.06 (dd, J=9.5 and 7.5 Hz, 1H, H-2), 3.39 (m, 1H, H-3), 3.75 (s, 3H, CH₃O), 3.78 (s, 2H, NCH₂Ar), 6.78 (d, J = 8.5 Hz, 2H, 3'-H), 7.07 (d, J = 8.5 Hz, 2H, 2'-H), 7.25 (m, 1H), 7.30– 7.34 (m, 4H); ¹³C NMR (74.5 MHz, CDCl₃) δ 30.5 (C-6), 32.9 (C-10), 35.1 (CH₂Ar), 38.6 (C-7), 38.9 (C-9), 42.1 (C-4), 49.9 (NCH₂), 52.4 (NCH₂Ar), 54.4 (C-3), 55.2 (CH₃O), 57.2 (C-2), 61.9 (C-5), 113.6 (C-3'), 127.0 (p-C), 128.2 and 128.4 (o-C and m-C), 129.5 (C-2'), 132.4 (C-1'), 139.9 (*ipso*-C), 157.8 (C-4'), 211.2 (C-8). Anal. calcd for $C_{25}H_{32}N_2O_3.H_2O$: C, 73.14; H, 8.34; N, 6.82. Found: C, 73.41; H, 8.08; N, 6.94.

trans-3-Benzylamino-1-[2-(4-methoxyphenyl)ethyl]-1-aza-spiro[4.5]decan-8-ol (8). To a cooled (0°C) solution of ketone 7 (0.58 g, 1.5 mmol) in MeOH (20 mL) was added NaBH₄ (112 mg, 3 mmol). After 30 min at 0°C and 1.5 h at room temperature, the reaction mixture was

poured into saturated aqueous NH₄Cl and extracted with CH_2Cl_2 (4×25 mL). The organic extracts were dried and concentrated. Chromatography (CH₂Cl₂ to 88:12 CH₂Cl₂:MeOH) yielded alcohol **8** (0.44 g, 75%): ¹H NMR (300 MHz, CDCl₃) δ 1.20–1.33 (m, 4H, H-6 and H-9), 1.35 (br d, J = 13.5 Hz, 1H, H-10eq), 1.42 (td, J=13.5 and 3.2 Hz, 1H, H-10ax), 1.46 (dd, J=13 and 6 Hz, 1H, H-4), 1.57 (br s, 1H, NH), 1.83 (m, 2H, H-7), 2.11 (dd, J=13 and 8.7 Hz, 1H, H-4), 2.48 (m, 1H, CH₂N), 2.54–2.65 (m, 3H, CH₂N and CH₂Ar), 2.77 (dd, J=9.5 and 4.5 Hz, 1H, H-2), 2.94 (dd, J=9.5 and 7 Hz, 1H, H-2), 3.23 (m, 1H, H-3), 3.41 (m, 1H, H-8), 3.69 (s, 2H, NCH₂Ar), 3.71 (s, 3H, CH₃O), 6.74 (d, J = 8.5 Hz, 2H, 3'-H), 7.04 (d, J = 8.5 Hz, 2H, 2'-H), 7.18 (m, 1H), 7.24–7.27 (m, 4H); ¹³C NMR (74.5 MHz, CDCl₃) δ 28.8 (C-6) 31.6 (C-10), 32.9 (C-9), 33.1 (C-7), 35.3 (CH₂Ar), 42.7 (C-4), 50.1 (NCH₂), 52.4 (NCH₂Ar), 54.5 (C-3), 55.2 (CH₃O), 57.2 (C-2), 62.6 (C-5), 70.6 (C-8), 113.6 (C-3'), 126.9 (p-C), 128.2 and 128.4 (o-C and m-C), 129.6 (C-2'), 132.8 (C-1'), 140.2 (*ipso-C*), 157.8 (C-4').

trans-3-[N-Benzyl-N-(tert-butoxycarbonyl)amino]-1-[2-(4-methoxyphenyl)-ethyl]-1-azaspiro[4.5]decan-8-ol (9). To a solution of alcohol 8 (430 mg, 1.1 mmol) in 9:1 MeOH:Et₃N (7 mL) was added di*tert*-butyl dicarbonate (476 mg, 2.2 mmol), and the mixture was heated at 50°C for 5h. The solvent was removed in vacuo, and the residue was purified by chromatography (CH₂Cl₂ to 88:12 CH₂Cl₂:MeOH) to give carbamate **9** (0.41 g, 76%): IR (CHCl₃) 1681; ¹H NMR (300 MHz, CDCl₃) δ 1.15–1.75 (m, 17H), 1.80–1.96 (m, 2H), 2.22–2.44 (m, 2H), 2.45–2.64 (m, 2H), 2.72 (m, 1H), 2.82 (m, 1H), 2.94 (m, 1H), 3.46 (m, 1H, H-8), 3.78 (s, 3H, CH₃O), 4.43 (br s, 2H, NCH₂Ar), 4.85 (br s, 1H, OH), 6.78 (d, J=8.7 Hz, 2H, 3'-H), 7.04 (d, J=8.7 Hz, 2H, 2'-H),7.13–7.34 (m, 5H); ¹³C NMR (74.5 MHz, CDCl₃) δ 26.1 (CH₂), 28.2 (CH₃), 32.6 (CH₂), 32.9 (CH₂), 34.8 (CH₂Ar), 39.5 (C-4), 46.6 (NCH₂Ar), 49.6 (NCH₂), 52.3 (C-3), 54.1 (C-2), 55.1 (CH₃O), 62.8 (C-5), 70.3 (C-8), 79.7 (C), 113.5 (C-3'), 126.3 (p-C), 125.9 and 128.1 (o-C) and m-C), 129.4 (C-2'), 132.4 (C-1'), 140.2 (ipso-C), 155.9 (CO), 157.7 (C-4'). Anal. calcd for $C_{30}H_{42}N_2$ O₄.H₂O: C, 70.28; H, 8.65; N, 5.46. Found: C, 70.37; H, 8.33; N, 5.46.

trans-3-[N-Benzyl-N-(tert-butoxycarbonyl)amino]-1-[2-(4-methoxyphenyl)ethyl]-1-azaspiro[4.5]dec-8-yl dibenzyl phosphate (10). To a solution of alcohol 9 (370 mg, 0.75 mmol) in CH₂Cl₂ (37 mL) were added 1-H-tetrazole (158 mg, 2.25 mmol) and N,N-diisopropyl dibenzyl phosphoramidite (0.38 mL, 1.12 mmol). The mixture was stirred at room temperature for 5h and then cooled to -50° C. MCPBA (98%, 197 mg, 1.12 mmol) in CH₂Cl₂ (10 mL) was added. The resulting solution was stirred at 0°C for 2h, diluted with CH₂Cl₂ and washed with saturated aqueous NaHCO₃ ($4\times50\,\mathrm{mL}$). The organic extracts were dried and concentrated. Chromatography (CH₂Cl₂ to 95:5 CH₂Cl₂:MeOH) yielded phosphate **10** (0.41 g, 73%): IR (CHCl₃) 1681; ¹H NMR (300 MHz, CDCl₃) δ 1.10–1.54 (m, 16H), 1.93 (m, 2H), 2.25 (m, 1H), 2.33 (m, 1H), 2.54 (m, 2H), 2.65 (m, 1H), 2.79 (m, 1H), 2.92 (m, 1H), 3.78 (s, 3H, CH₃O), 4.10 (br s, 1H), 4.40 (br s, 2H, NCH₂Ar), 4.70 (br s, 1H), 4.98 (s, 2H, OCH₂Ar), 5.01 (s, 2H, OCH₂Ar), 6.78 (d, J=8.7 Hz, 2H, 3'-H), 7.01 (d, J=8.7 Hz, 2H, 2'-H), 7.10–7.40 (m, 15H); ¹³C NMR (74.5 MHz, CDCl₃) δ 25.9 (CH₂), 28.2 (CH₃), 30.5 (CH₂), 31.0 (CH₂), 32.8 (CH₂), 34.9 (CH₂Ar), 39.7 (C-4), 46.6 (NCH₂Ar), 49.4 (NCH₂), 52.3 (C-3), 54.1 (C-2), 55.1 (CH₃O), 62.0 (C-5), 68.9 and 69.0 (OCH₂Ar), 77.5 (C-8), 79.7 (C), 113.5 (C-3'), 126.3 (*p*-C), 125.9 and 128.1 (*o*-C and *m*-C), 127.8 (CH), 128.3 and 128.4 (CH), 129.4 (C-2'), 132.5 (C-1'), 135.8 and 135.9 (C), 140.3 (*ipso*-C), 155.9 (CO), 157.7 (C-4'); ³¹P NMR (121.5 MHz, CDCl₃) δ –5.44. Anal. calcd for C₄₄H₅₅N₂O₇P.1/8H₂O: C, 69.80; H, 7.36; N, 3.70. Found: C, 69.40; H, 7.31; N, 3.73.

In some runs, the corresponding *N*-oxide was formed as a minor byproduct:

trans-3-[N-Benzyl-N-(tert-butoxycarbonyl)amino]-8-(dibenzylphosphoroxy)-1-[2-(4-methoxyphenyl)ethyl]-1-azaspirol4.5|decane 1-oxide. ¹H NMR (300 MHz, CDCl₃) δ 1.20–1.65 (m, 15H), 1.90–2.10 (m, 2H), 2.28 (m, 1H), 2.32–2.48 (m, 2H), 2.87 (m, 1H), 3.13 (m, 2H), 3.44–3.62 (m, 2H), 3.78 (s, 3H, CH₃O), 3.88 (m, 1H), 4.05 (m, 1H), 4.80-5.20 (m, 6H), 6.83 (d, J=8.6 Hz, 2H, 3'-H), 7.13 (d, J = 8.6 Hz, 2H, 2'-H), 7.18–7.37 (m, 15H); ¹³C NMR (74.5 MHz, CDCl₃) δ 28.1 (CH₃), 29.3 (CH₂), 30.2 (CH₂), 30.3 (CH₂), 33.8 (CH₂), 46.5 (NCH₂Ar), 50.4 (C-3), 55.2 (CH₃O), 61.7 (CH₂N), 68.9 and 69.0 (C-5), 69.2 and 69.3 (OCH₂Ar), 69.8 (C-2), 76.0 and 76.1 (C-8), 80.4 and 81.3 (C), 114.1 (C-3'), 126.3 (p-C), 125.9 and 128.2 (o-C and m-C), 127.9 (CH), 128.5 and 128.6 (CH), 129.8 (C-2'), 135.6 and 135.7 (C), 140.6 (ipso-C), 156.5 (CO), 158.3 (C-4').

trans-3-[N-Benzyl-N-(tert-butoxycarbonyl)amino]-1-[2-(4-methoxyphenyl)ethyl]-1-azaspiro[4.5]dec-8-yl dihydrogenphosphate (11). A mixture of phosphate 10 (280 mg, 0.37 mmol) and 10% Pd/C (140 mg) in MeOH (40 mL) was stirred under 1 atm H₂ for 12h. The suspension was filtered through Celite, and the filtrate concentrated to give phosphate 11 (195 mg, 92%): IR (film) 3420, 1690, 1661; ¹H NMR (200 MHz, CD₃OD) δ 1.10–2.20 (m, 21H), 2.42 (br s, 1H), 2.80 (br s, 2H), 3.32 (m, 1H), 3.67 (s, 3H), 3.85 (br s, 1H), 4.32 (br s, 1H), 4.38 (s, 2H), 6.76 (d, J = 8.4 Hz, 2H), 6.98–7.35 (m, 7H); ¹³C NMR (74.5 MHz, CD₃CN) δ 25.6 (CH₂), 28.5 (CH₃), 30.0 (CH₂), 31.2 (CH₂), 36.5 (CH₂), 38.2 (CH₂), 50.1 (CH₂), 51.0 (CH₂), 53.7 (CH), 55.9 (CH₃), 73.1 (CH), 74.1 (C), 81.4 and 82.0 (C), 115.0 (CH), 127.8 (CH), 129.4 (CH), 131.1 (CH), 139.9 (C), 156.9 (C), 159.6 (C). ISMS: 576 (M+H).

trans-3-(Benzylamino)-1-[2-(4-methoxyphenyl)ethyl]-1-azaspiro[4.5]dec-8-yl dihydrogenphosphate (2). To a solution of carbamate 11 (165 mg, 0.29 mmol) in CH₂Cl₂ (13 mL) was added at 0°C TFA (3.3 mL, 43 mmol). After 12 h at room temperature, the solvent and the excess of acid were removed in vacuo to give the trifluoroacetic acid salt of 2 (170 mg, quantitative): IR (film) 2800 (br), 1668; ¹H NMR (200 MHz, CD₃OD) δ 1.40−2.30 (m, 10H), 2.80−3.20 (m, 3H), 3.40 (m, 1H), 3.67 (s, 3H), 3.60−4.00 (m, 2H), 4.00−4.30 (br, 4H), 6.79 (d, J=8.8 Hz, 2H), 7.13 (d, J=8.8 Hz, 2H), 7.30−7.50

(m, 5H); 13 C NMR (74.5 MHz, DMSO- d_6) δ 29.3 (CH₂), 29.7 (CH₂), 30.3 (CH₂), 30.7 (CH₂), 35.8 (CH₂), 48.8 (CH₂), 49.3 (CH₂), 52.1 (CH), 55.3 (CH₃), 60.5 (CH₂), 70.9 (C), 72.3 (CH), 114.2 (CH), 117.1 (q, C) 129.0 (CH), 129.1 (CH), 129.3 (CH), 130.1 (CH), 132.1 (C), 158.4 (C), 159.1 (q, C). ISMS: 475 (M+H). This trifluoroacetate (50 mg) was dissolved in MeOH (12 mL) containing K₂CO₃ (30 mg). The solvent was removed to give the dipotassium salt of 2: ¹H NMR (300 MHz, CD₃OD) 8 1.26-1.66 (m, 8H), 2.13 (br, 2H), 2.23 (dd, J = 13 and 8.6 Hz, 1H), 2.56 (m, 1H), 2.60–2.72 (m, 3H), 2.78 (dd, J=9.5 and 5.7 Hz, 1H), 3.01 (dd, J=9.5 and 7.4 Hz, 1H), 3.25 (m, 1H), 3.70–3.75 (m, 5H), 3.90 (br, 1H), 6.79 (d, J = 8.7 Hz, 2H), 7.08 (d, J = 8.7 Hz, 2H), 7.20–7.40 (m, 5H); 13 C NMR (74.5 MHz, CD₃OD) δ 30.5 (CH₂), 32.6 (CH₂), 32.9 (CH₂), 32.9 (CH₂), 36.0 (CH₂Ar), 43.1 (C-4), 51.4 (NCH₂), 53.1 (NCH₂Ar), 55.3 (OMe), 55.7 (C-3), 57.8 (C-2), 64.3 (C-5), 73.7 (d, $J_{\rm CP} = 4.5 \, \text{Hz}$, C-8), 114.7 (C-3'), 128.2 (p-C), 129.5 and 129.6 (o-C and m-C), 130.6 (C-2'), 133.8 (C-1'), 140.2 (*ipso-C*), 157.8 (C-4').

Acknowledgments

Support for this research was provided by DGES (Spain) through Grant PB97-0877. Thanks are also due to the 'Comissionat per a Universitats i Recerca' (Generalitat de Catalunya) for Grant SGR97-00166 and to the MEC (Spain) for a fellowship to G.P.

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